



**Jordan University of Science and Technology**  
**Faculty of Applied Medical Sciences**  
**Department of Medical Laboratory Sciences**  
**Course Syllabus**

<b>Course Information</b>	
<b>Course Title</b>	Diagnostic Immunology and Serology (2 credit hours)
<b>Course Code</b>	LM 335
<b>Prerequisites</b>	LM 232
<b>Course Website</b>	<a href="http://www.just.edu.jo">http://www.just.edu.jo</a>
<b>Instructor</b>	Dr. Muhamad Shakhatreh
<b>Office Location</b>	M5L-4; Room no. 2
<b>Office Phone #</b>	23874
<b>Office Hours</b>	To be determined
<b>E-mail</b>	mkshakhatreh@just.edu.jo
<b>Course Description</b>	
<p>This course introduces the concepts of clinical immunology and serology for clinical laboratory practice. It covers essential theoretical principles along with serology techniques most commonly used in the lab. It provides students with knowledge required to perform different serological techniques used in disease diagnosis. It consists of the theory, application, and performance of common serological testing used in a clinical lab including agglutination reactions, precipitation reactions, complement fixation test (CFT), direct and indirect hemagglutination (HA and IHA), hemagglutination inhibition (HAI), Radioimmunoassay (RIA) including instrumental production of immune serum, labeling of antigen, commercial kits, immunodiffusion, immunoelectrophoresis, direct and indirect fluoroimmunoassays (FIA), Enzyme-linked immunosorbent assay (ELISA), and immunoblotting. The course is accompanied by a practical laboratory applications course (LM 337)</p>	

<b>Textbook</b>	
<b>Title</b>	Manual of Clinical Laboratory Immunology
<b>Author(s)</b>	Barbara Deetrick
<b>Publisher</b>	American society of microbiology
<b>Year</b>	2002
<b>Edition</b>	6th
<b>Book Website</b>	
<b>Other references</b>	Medical immunology, internet clinical immunology and serology resources.

<b>Assessment</b>		
<b>Assessment</b>	<b>Expected Due Date</b>	<b>Percentage</b>
<b>First Exam</b>	Week 5	<b>30%</b>
<b>Second Exam</b>	Week 10	<b>30%</b>
<b>Final Exam</b>	To be determined	<b>40%</b>

<b>Course Objectives</b>	<b>Percentage</b>
1. To provide students with basic principles of antigen antibody reactions, precipitation, agglutination, complement fixation (CF), Radioimmunoassay (RIA), ELISA, and fluorescent antibody technology.	30%
2. To introduce students to applications of immunological techniques in the diagnosis of bacterial infections, viral infections, parasitic infections, as well as immunological diseases	50%
3. To introduce students to principles and techniques for laboratory assessment of host immunity	20%

<b>Teaching &amp; Learning Methods</b>
Lectures, PowerPoint Presentations.

<b>Learning Outcomes: Upon successful completion of this course, students will be able to:</b>	
<b>Related Objective(s)</b>	<b>Reference(s)</b>
1. Describe the principles of Ag-Ab reactions and their usage in clinical laboratories.	
2. Perform sero-identification of bacteria, Widal test, Brucella test, blood grouping, cold agglutinins, and Paul Bunnell test.	
3. Perform CRP, RA, Pregnancy test, Meningo test, IHA for hydatid disease, and HbsAg.	
4. Utilize precipitation tests to do Lancefield grouping of Streptococci, double diffusion, Immunoelectrophoresis and immunofixation, Radial immunodiffusion (RID) to quantitate Ig, C3, C4, Haptoglobin, etc.	
5. Use complement fixation test (CFT) for antibody detection of viruses, bacteria, and parasites, as well as tissue typing.	
6. Utilization of Radioimmunoassay RIA and ELISA technology to detect hepatitis markers, total IgE, allergen specific IgE, BHCG, TORCH, etc.	
7. Use direct immunofluorescence (DIF) for detection of Chlamydia, Treponema, Viruses, and immune complexes in tissues.	

8. Use immunofluorescence to detect anti-nuclear antibodies (ANA), anti-mitochondrial antibodies (AMA), ASMA, anti-dsDNA, anti-parietal Cell Antibody APCA, anti-neutrophil cytoplasmic antibody (ANCA), and antibodies to parasites.	
9. Perform E-rosette count, B-cell count, lymphocyte transformation test (LTT), and evaluate skin test results.	

Course Content		
Week	Topics	Chapter in Textbook (handouts)
1	1-Agglutination reactions: A:Direct agglutination reactions: - Serological identification of bacteria. - Widal test.	
2	- Brucella agglutination test. - Blood grouping- ABO-Rh test. - Cold agglutinins and Heterophile antibodies. B:Passive agglutination or indirect agglutination. - C-reactive protein tests. - RA-latex test - Meningo-latex test.	
3	C:Passive haemagglutination tests: - Hepatitis B surface Antigen(HbsAg). - Ecchinococcus granulosus antibodies(Hydatid cyst). - Monospot test and Paul-Bunnell test.	
4	2- Precipitation reactions: A: Tube precipitation reactions: - Ascoli test –Anthrax. - Lancefield grouping of Streptococcus. B-:Gel precipitation reactions: - Double diffusion – Antigen identification and relation.	
5	<b>Exam 1</b> - Radial immunodiffusion: Immunoglobulins, Complement	
6	Ceruloplasmin, Haptoglobulins, Transferin. Immuno-electrophoresis and Immunofixation	
7	3- Complement fixation tests: - Antibodies to viruses, bacterial antigens, parasites.	
8	- Microcytotoxicity tests. 4-Neutralization reactions: - Anti Streptolysin O test. - Virus infectivity tests.	

9-10	<p>5-Radioimmunoassay :</p> <ul style="list-style-type: none"> <li>- Hepatitis markers</li> <li>- Total serum IgE-Prest.</li> <li>- Allergen specific IgE – Radio-Allergo-Sorbent Test (RAST).</li> <li>- BHCG.</li> </ul> <p><b>2<sup>nd</sup> Exam</b></p>	
11-12	<p>6- Enzyme-linked immunosorbent assay(ELISA):</p> <ul style="list-style-type: none"> <li>- TORCH test.</li> <li>- HIV test.</li> <li>- BHCG</li> <li>- Total and specific IgE.</li> </ul>	
13-14	<p>7-Flourescent Antibody technology:</p> <ul style="list-style-type: none"> <li>- Detection of Antigens: Chlamydia, Treponema.</li> <li>- Detection of immune complexes in tissues.</li> <li>- Detection of Antibodies: ANA, AMA, ASMA, AntiDNA, APCA, ANCA</li> <li>- Detection of Ab to Toxoplasma, Trichinella, Amoeba.</li> </ul>	
15	<p>8-Assessment of Cellular Immunity:</p> <ul style="list-style-type: none"> <li>- T-cell count – E-Rosette.</li> <li>- B-Cell count.</li> <li>- T-cell function: Lymphocyte transformation test.</li> <li>- Skin tests</li> </ul> <p>New methods used in clinical immunology and serology</p>	
16	<b>Final Exam</b>	

<b>Additional Notes</b>	
<p><b><u>Attendance policy:</u></b> Mandatory for all of the lectures. Students are expected to attend more than 80% of lectures.</p> <p><b><u>Expected workload:</u></b> Students are expected to attend all classes, and pass the exams.</p> <p><b><u>Feedback:</u></b> Feedback or concerns from the students regarding the progression in the course and the material covered in the lecture can be discussed with the instructor at designated office hours or by appointment.</p>	

The Clinical Laboratories<sup>™</sup> Regulation places an emphasis on facility design and services criteria with a focus on quality of services and safety of professionals based on the local and federal laws in addition to international accreditation standards. Therefore, this document provides a base for the Health Regulation Department (HRD) to assess the Clinical Laboratories performance in Dubai and to ensure a safe and competent delivery of services. It will also assist Clinical Laboratories in developing their quality management systems and in assessing their own competence to ensure compliance with Clinical Immunology publishes original research on the molecular and cellular bases of immunological disease. The journal also features reviews of timely topics in basic immunology, case reports, and letters to the editor. Research Areas Include

- Ageing
- Allergy
- Autoimmunity
- Biotechnology
- Clinical Laboratory Immunology
- HIV
- Immunodermatology
- Immunohematology
- Immunotoxicology/Environmental
- Infection and Immunity
- Mucosal Immunity
- Neuroimmunology
- Primary Immunodeficiency
- Transplantation
- Tumor Immunology
- Vaccines.

Benefits to authors We also provide many author benefits, such as free PDFs, a liberal copyright policy, special discounts on Elsevier publications and much more.

Xxv, 1322 pages : 29 cm. Includes bibliographical references and indexes. General methods -- General methods: antibody-based and molecular -- Immunoglobulin methods -- Complement -- Flow cytometric analysis -- Cellular immunology assays -- Cytokines, chemokines, and adhesion molecules -- Immunohistology and immunopathology -- Immunologic diagnosis and monitoring of disease -- Infectious diseases caused by bacteria, mycoplasmas, chlamydiae, and rickettsiae -- Mycotic and parasitic. Diseases -- Viral diseases -- Human immunodeficiency virus -- Immunodeficiency diseases -- Allergic diseases -- Sys , North Carolina Clinical Immunology and Serology: A Laboratory Perspective, Third Edition Christi Molecular Cloning: A Laboratory Manual. 2,231 PagesÂ·2010Â·123.37 MBÂ·5,439 DownloadsÂ·New! Molecular Cloning: A Laboratory Manual Joseph Sambrook|David W. Russel Henryâ€™s Clinical Diagnosis and Management by Laboratory Methods, 23e. 1,823 PagesÂ·2016Â·193.93 MBÂ·35,269 DownloadsÂ·New!Â Molecular Cloning: A Laboratory Manual, Third Edition Tietz Fundamentals of Clinical Chemistry and Molecular Diagnostics. 1,103 PagesÂ·2014Â·198.75 MBÂ·18,709 DownloadsÂ·New! Chemistry and Molecular Diagnostics, Tietz Fundamentals of Clinical Chemistry and Molecular Diagno Essential Clinical Immunology (PDF) - SACEMA. In clinical laboratories, fluorescent antibody tests are currently used for detection of bacterial, viral, and fungal infections as well as for bioimaging of tissue samples. A number of respiratory viruses can be directly detected in nasopharyngeal samples using direct fluorescent antibody (DFA) test.Â In: Rose NR, Hamilton RG, Detrick B, eds. Manual of Clinical Laboratory Immunology. Washington, DC: ASM Press; 2002:6-25. 6. Ridker PM, Morrow DA. Start by marking ["Manual of Clinical Laboratory Immunology"](#) as Want to Read: Want to Read savingâ€¦| Want to Read. Currently Reading. Read. Manual of Clinical Lab by Noel R. Rose. Other editions.Â The much-awaited fifth edition of this classic reference manual features the latest on standard and cutting-edge immunological tests and procedures in clinical laboratory immunology. The best features of earlier editions have remained in this update, including a step-by-step interpretive approach to methodology, with emphasis on clinical applications and interpretation. The much-awaited fifth edition of this classic reference manual features the latest on standard and cutting-edge immunological tests and procedures in clinical laboratory immunology. Essential Clinical Immunology begins with the basic concepts and then details the immunological aspects of various disease states involving major organs of the body. The book explores how we can better understand disease and its treatment through clinical immunology. Looking forward, each chapter concludes with patterns for future research. John B. Zabriskie (M.D., Columbia College of Physicians and Surgeons) is Professor Emeritus and former head of the Laboratory of Clinical Microbiology and Immunology at The Rockefeller University, New York, New York.Â Manual of Clinical Laboratory Immunology. Washington, DC: American Society for Microbiology; 1997. von Muhlen CA, Tan EM.